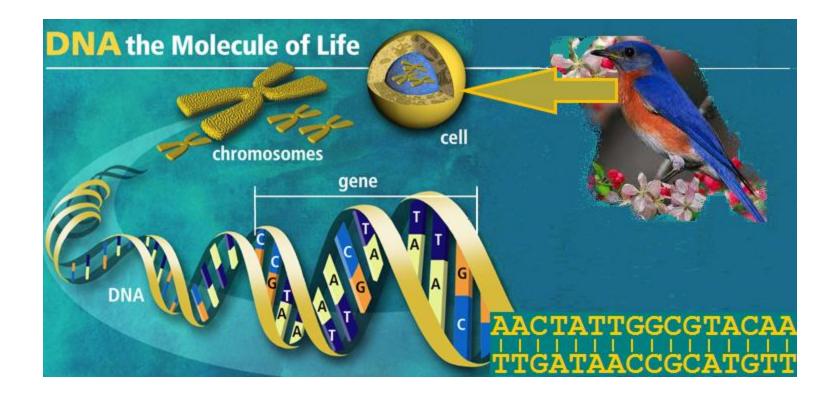
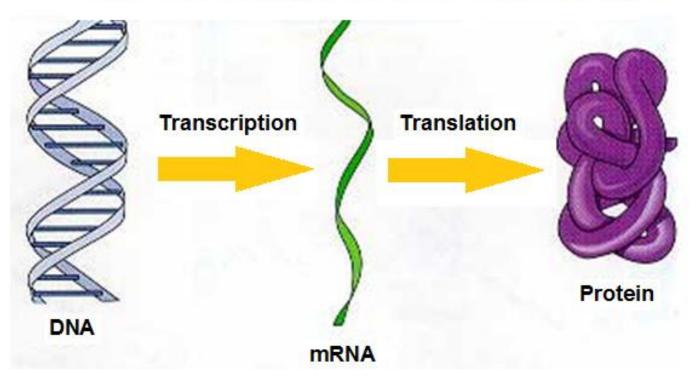
Mapping Reads to Reference Genome



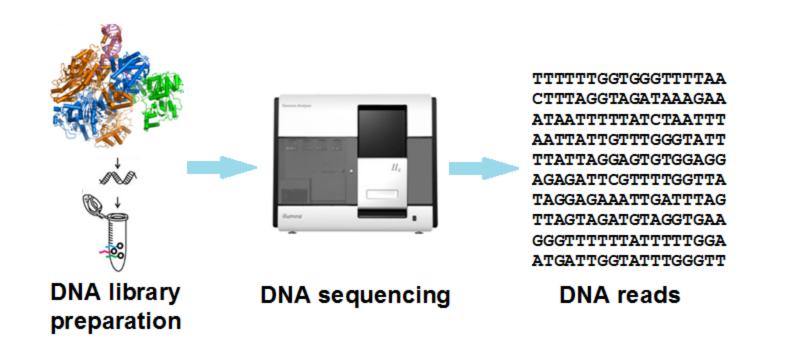
- DNA carries genetic information
- DNA is a double helix of two complementary strands formed by four nucleotides (bases):

Adenine, Cytosine, Guanine and Thymine

The Central Dogma of Molecular Biology



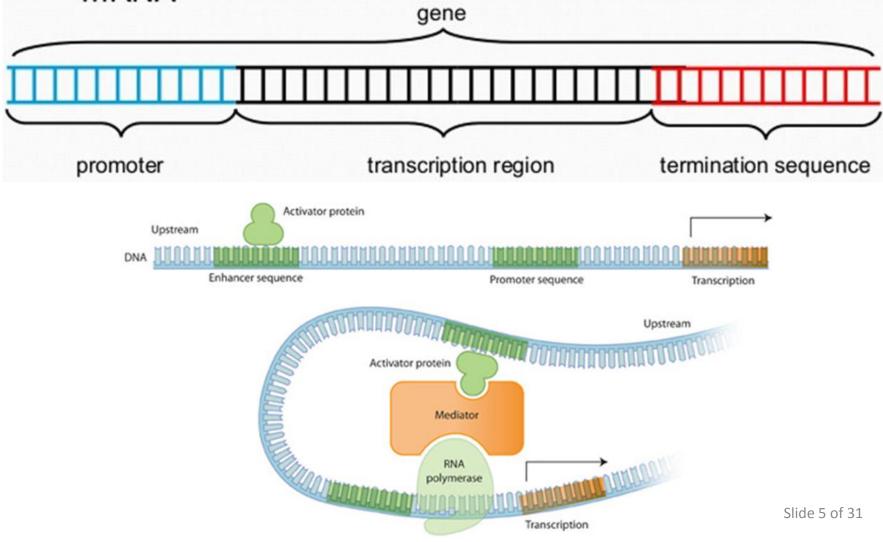
- Gene expression is the process by which DNA is transcribed into mRNA (eventually translated into proteins)
- Mechanisms controlling gene expression are not fully understood yet



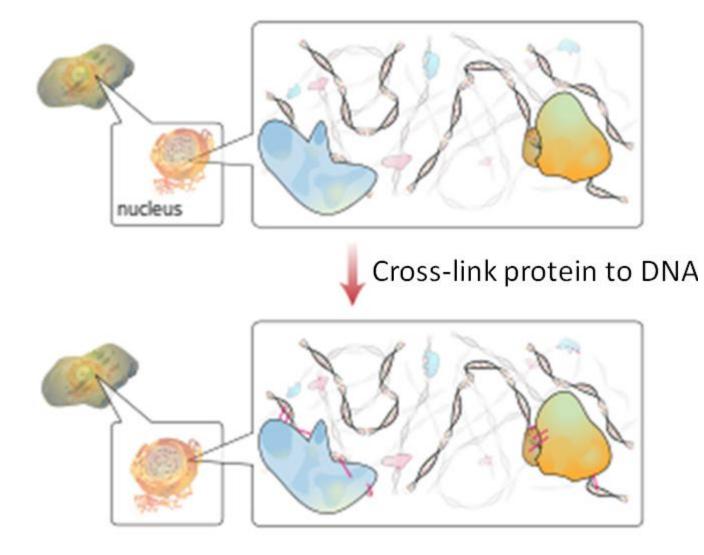
- New-generation sequencing technology allows fast and inexpensive DNA sequencing
- Helps biologists to study cellular processes

Example: Identify Transcription Factors binding sites

Promoters are located upstream from the DNA region that contains the information to be transcribed into mRNA

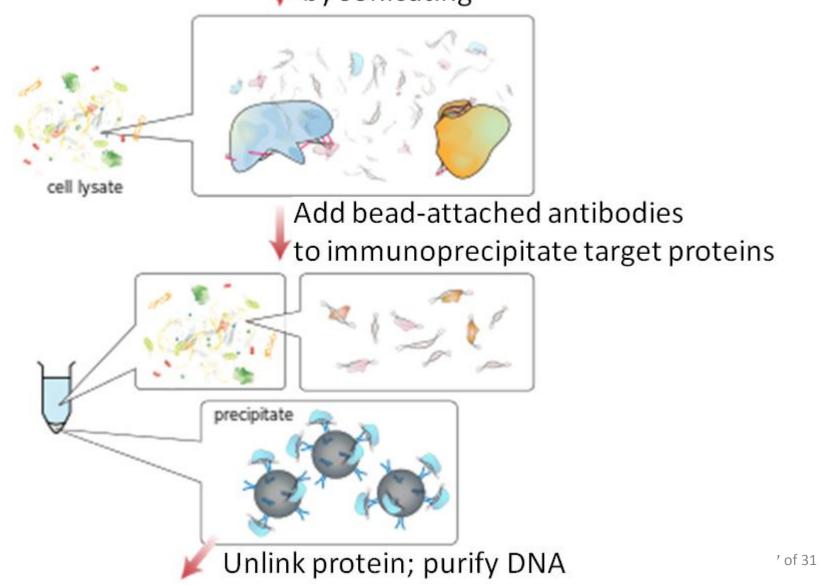


Example: Identify Transcription Factors binding sites

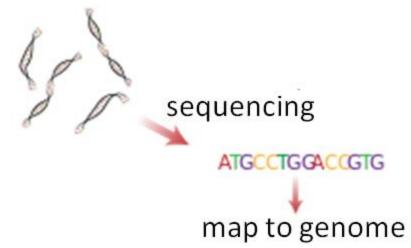


Cell diagram adapted from LadyOfHats' Animal Cell diagram. Wikipedia.

Example: Identify Transcription Factors binding sites Shear DNA strands by sonicating



Example: Identify Transcription Factors binding sites



Reference genome

TAGAAATTGAAACAGCTGTGTTTAGTGCCTTTGTTCA----ACCCCCTTGCAACAACCTTGAGAACCCCCAGGGAATTTGT TATATT ATGCTATTCAGTTCTAAATATAGAAATTGAAACAG GTGTTTAGTGCCTTTGTTCA----ACCCCCTTGCAACAAC aaccccagggaatttgt acagetgtgtttagtgeetttgttea----acceeettg aacaacettgagaaceecagggaatttgt tatatttatgetatteagttetaaatatagaaatt TATAT TATGCTATTCAGTTCTAAATATAGAAATTGAAAACA etqtqtttaqtqcetttqttca----acceeettqcaac ACCTTGAGAACCCCAGGGAATTTGT TATATTTA getatteagttetaaatatagaaattgaaacaget GTTTAGTGCCTTTGTTCACATAGACCCCCTTGCAA aacettgagaaceeeaqqqaatttqt TATATTTATGCTATTCAGT GAAATTGAAACAGCTGTGTTTAGTGCCTTTGTTCA ccccttacaacaaccttgagaaccccagggaattt tatatttatgetatteagt GCCTTTGTTCACATAGACCCCCTTGCAACAACCTT cagggaatttgt tatatttatgetatteagtteta AG----ACCCCCTTGCAACAACCTTGAGAACCCCCAGGGA TATATTTATGCTATTCAGTTCTAA A----ACCCCCTTGCAACAACCTTGAGAACCCCCAGGGAA TATATTTATGCTATTCAGTTCTAAA A----ACCCCCTTGCAACAACCTTGAGAACCCCCAGGGAA TATATTTATGCTATTCAGTTCTAAA TGCAACAACCTTGAGAACCCCAGGGAATTTG7 TATATTTATGCTATTCAGTTCTAAAT TGCAACAACCTTGAGAACCCCAGGGAATTTG1 TATATTTATGCTATTCAGTTCTAAAT TGCAACAACCTTGAGAACCCCCAGGGAATTTGT tatatttatgetatteagttetaaatatagaaatt tgcaacaacettgagaaceccagggaatttgt tatatttatgetatteagttetaaatatagaaatt CAACCTTGAGAACCCCAGGGAATTTGT TATTTATECTATTCAGTTATAAATATAGAAATTGAAACAG CCTTGAGAACCCCAGGGAATTTGT atttatgetatteagttetaaatatagaaattgaa CTTGAGAACCCCAGGGAATTTGT tttacgctattcagtactaaatatagaaattgaaa CTTGAGAACCCCAGGGAATTTG1 ttatgctattcagttctaaatatagaaattgaaac gggaatttgt reads

40.000

60.000

20.000

100,000

80,000

- Mapping DNA reads back to a reference genome is the first step in the data analysis
- Mapping short sequenced reads back to a reference genome is a string search problem: given a text and a query, find all (approximate) occurrences of the query in the text



Group Work

- Assume that a human reference genome is given (a string of 3 billion characters long)
- Assume that you need to map 1 million 50bp reads to the genome
- Come up with a method to map fast the reads to the genome

Problem statement:

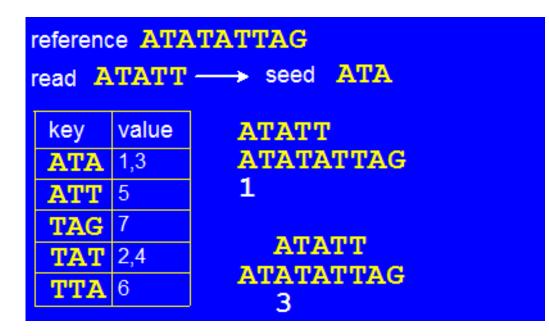
Given a string S of length *n* and a short string P of length *m* (n >> m), find all locations where P occurs in S

ATATGTTAGTCAAGTTAAGACCTATGTTAG

Methods for Mapping Short Reads

- To speed up mapping, search space is reduced by focusing only on those regions of genome that share the same seed(s) with a read
- A seed, or k-mer (q-gram), is a substring of a read of length k
- Common data structures to index the data (genome) and speed-up the search:
 - hash tables
 - \succ suffix trees
 - suffix arrays
 - Burrows-Wheeler transform (BWT) with Ferragina-Manzini (FM) index

Hash Indexing



- Hash all genome k-mers into a hash table using seeds of fixed length k as hash keys, and corresponding genomic positions as values
- Use the *k*-mers in a read as hash keys to retrieve locations that are potential hits
- Align the entire read to the potential locations and count the number of mismatches

Hash Indexing

| Seed Size | Table Size | Space, GB |
|-----------|----------------------------|-----------------|
| 32 | 18,446,744,073,709,600,000 | 147,573,952,590 |
| 28 | 72,057,594,037,927,900 | 576,460,752 |
| 24 | 281,474,976,710,656 | 2,251,800 |
| 20 | 1,099,511,627,776 | 8,796 |
| 18 | 68,719,476,736 | 550 |
| 16 | 4,294,967,296 | 34 |
| 12 | 16,777,216 | 134 MB |

Hash Indexing

Disadvantages:

- 1. The longer seeds, the more space demanding
- 2. The shorter seeds, the more time consuming

Group Work

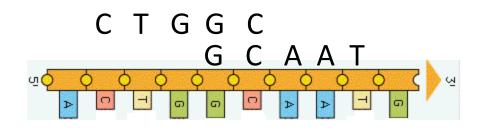
1. Build a hash table for the following sequence using seeds of length 2 and 3

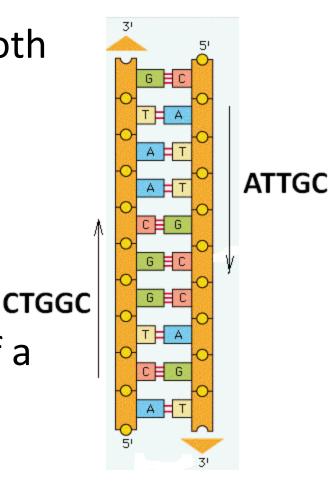
ATATGTTAGTCAAGTTAAGACCTATGTTAG

2. Map read TATG to the given sequence using the seed TA (TAT) and your hash tables. How many different alignments did you have to make?

Mapping

- 1. Reads are generated from both strands of DNA
- 2. Reads are always sequenced from 5' to 3'
- Mapping is performed to only (+) strand of DNA
- 4. Map reverse-complement of a read: ATTGC, rc: GCAAT



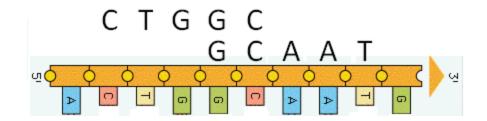


Mapping

Output format

Read_ID Read Chromosome Position Strand

- 1 CTGGC 1 1 +
- 2 ATTGC 1 4 -



Encoding of Reads

- A = 00
- C = 01
- G = 10
- T = 11
- ATTGC = 0011111001

Advantages:

- 1. Each character takes 2 bits instead of 8 bits
- 2. Retrieval of all positions where a seed occurs takes O(1) time (use encoding of a seed as an index for a hash table's bin)

Suffix Array

| reference ATATATTAG> ATATATTAG\$ read ATATT> seed ATA | | | |
|--|--|--|--|
| exicographic sorting | | | |
| \$ATATATTA <mark>C</mark> | | | |
| AG\$ATATATT | | | |
| ATATATTAG\$ - ATA | | | |
| ATATTAG\$AT 🔶 🗖 🗖 🗖 | | | |
| ATTAG\$ATA <mark>T</mark> | | | |
| G\$ATATATT <mark>A</mark> | | | |
| TAG\$ATATAT | | | |
| TATATTAG\$A | | | |
| TATTAG\$AT <mark>A</mark> | | | |
| TTAG\$ATAT <mark>A</mark> | | | |
| | | | |

- Find all circular shifts of the reference genome
- Lexicographically sort the circular shifts
- All circular shifts that start with the same substring are consecutive
- Record the starting indices of the circular shifts

12345678910 ATATATTAG\$

10 8 1 3 5 9 7 2 4 6

Suffix Array

For a given string S, Pos[i] = j, such that S[j...n] is a prefix of row i in M

To find a given pattern W of length m, we know that all rows having W as a prefix in M are contiguous; hence, positions of P in S are stored in contiguous range [L, R] in the suffix array Pos

Μ

- 1 \$ATATATTAG
- 2 AG\$ATATATT
- 3 ATATATTAG\$
- 4 ATATTAG\$AT
- 5 ATTAG\$ATAT
- 6 G\$ATATATTA
- 7 TAG\$ATATAT
- 8 TATATTAG\$A
- 9 TATTAG\$ATA
- 10TTAG\$ATATA

Suffix Array

12345678910 ATATATTAG\$

10 8 1 3 5 9 7 2 4 6

М

- 1 \$ATATATTAG
- 2 AG\$ATATATT
- 3 ATATATTAG\$
- 4 ATATTAG\$AT
- 5 ATTAG\$ATAT
- 6 G\$ATATATTA
- 7 TAG\$ATATAT
- 8 TATATTAG\$A
- 9 TATTAG\$ATA

10TTAG\$ATATA

if
$$W \leq_P A_{Pos[0]}$$
 then
 $L_W \leftarrow 0$
else if $W \geq_P A_{Pos[N-1]}$ then
 $L_W \leftarrow N$
else
{ $(L, R) \leftarrow (0, N-1)$
while $R-L \geq 1$ do
{ $M \leftarrow (L+R)/2$
if $W \leq_P A_{Pos[M]}$ then
 $R \leftarrow M$
else
 $L \leftarrow M$
}
}

Suffix arrays: A new method for on-line string searches Udi Manber Gene Myers

12345678910 ATATATTAG\$



М

1 \$ATATATTAG

- 2 AG\$ATATATT
- 3 ATATATTAG\$
- 4 ATATTAG\$AT
- 5 ATTAG\$ATAT
- 6 G\$ATATATTA
- 7 TAG\$ATATAT
- 8 TATATTAG\$A
- 9 TATTAG\$ATA 10TTAG\$ATATA

Time to find all occurrences of W in S is O(|W|*log*(n)), where n = |S| Space to store a suffix array is 4n

The authors also proposed algorithm with time O(|W| + log(n))

Suffix arrays: A new method for on-line string searches Udi Manber Gene Myers

Input Format

Reference genome is usually given as a set of files, each file per chromosome.

Each file is in **Fasta** format:

>Region_2 taxid=9606|spec=Homo sapiens|chr=1|ctg=NC_000001|str=(-)|start=8014301|end=8014551|len=251
GCCCTGACATTTGAGGCGGCGCTGGTGCAAGGGGGGGAAATTCTGACACCGAGTTCTGTGAGGGGGCCCTGGGAACGGCTTC
GCCTCTGCGCCCACCAGTGTGAAGAGCCCCCCCCCAGCGTGGGGCCACCAGACACACTCCCACGGGGGCGCCGGG
AAACCTACCGACTGACCTGCATGACGTAGGTCCACTTCCGGCGGCGCCGCAGCATGCGCCACCCGGGATTGGCCGAGTCAGG
TCGCAGTGGGC

Input Format

Reads are usually given in **FASTQ** format:

@ERR030887.1 HWI-BRUNOP16X_0001:8:1:7336:1073#0/1 TNTCGATTACATGTGGATCAGGTTGATTTAATAATGGCGATAGGGNNNCT. @ERR030887.2 HWI-BRUNOP16X_0001:8:1:10288:1073#0/1 TNAGTCTTTCCCAGCCTAACAAAGAAAGCAAGAATAATTGGGCACNNNGA @ERR030887.4 HWI-BRUNOP16X_0001:8:1:15389:1074#0/1 @ERR030887.5 HWI-BRUNOP16X_0001:8:1:16693:1073#0/1 CNAGTCCGTCACTCCATCCTACCCTTATGGGCCAGGTAAGCCAACNNNCC

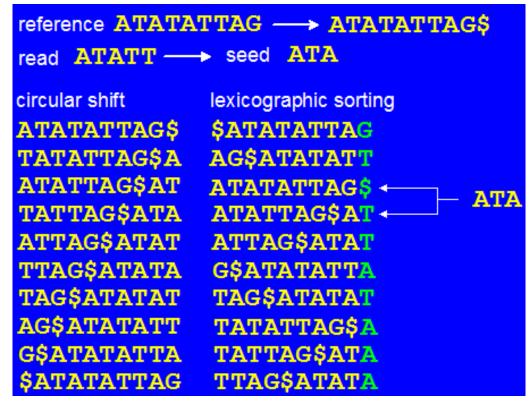
Read ID Sequenced Read Ignore Quality Info

Homework 1 discussion

Next Lecture

- Approximate string search
- Smith-Waterman algorithm
- Hash table, suffix array for approximate string search

Burrows-Wheeler Transform



- Build Burrows-Wheeler transform (BWT) for the reference genome
- Find positions within BWT corresponding to suffixes whose prefix is a seed of the read
- Calculate from these positions genomic positions
- Align the entire read to the potential locations and count the number of mismatches

Methods for Mapping Short Reads

| | Hash Indexing | Burrows-Wheeler transform |
|---|---------------|---------------------------|
| Seed length for a read | fixed | variable |
| Time to find genomic positions for a <i>k</i> -mer | O(1) | $O(k + Occ \cdot \log N)$ |
| Time to map entire read of length <i>P</i> to <i>Occ</i> genomic positions, where the <i>k</i> -mer occurs | O(Occ · P) | O(<i>Occ · P</i>) |

- The length of the seed used in hashing is fixed and usually shorter than the seed for BWT
- Hence, Occ with BWT is smaller than Occ with hash indexing
- We need to check a smaller number of full-length read alignments with BWT; thus, mapping of short reads with BWT is more time-efficient

Find positions within BWT corresponding to suffixes whose prefix is a seed of the read

1 \$ATATATTAG 2 AG\$ATATATT 3 ATATATTAG\$ 4 ATATTAG\$AT 5 ATTAG\$ATAT 6 G\$ATATATTA 7 TAG\$ATATTA 7 TAG\$ATATAT 8 TATATTAG\$A 9 TATTAG\$ATATA

F \$ 0 A 1 C 5 G 5 T 6 10

м

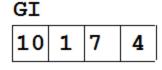
Given: P, a pattern of length p $BW_Search(P[1,p])$ c = P[p], i=p sp = F[c] + 1, ep = F[c+1]while sp < ep and i > 1 c = P[i-1], i = i - 1 sp = F[c] + Occ(c, 1, sp-1) + 1 ep = F[c] + Occ(c, 1, ep)print sp and ep

Example: $BW_Search(ATA)$ c = A, i = 3 sp = F[A] + 1 = 2, ep = F[A + 1] = F[C] = 5 i=3: c = T, sp = 6 + 0 + 1 = 7, ep = 6 + 3 = 9i=2: c = A, sp= 1 + 1 + 1 = 3, ep = 1 + 3 = 4

At each iteration *i*, sp points to the first row of M prefixed by P[i,p], and ep points to the last row of M prefixed by P[i,p]

Calculate Genomic Positions from [sp,ep]







F \$ 0 A 1 C 5 G 5 T 6 10

- Mark row of M corresponding to each 3-d genomic position
- Store explicitly these positions in array GI
- If i-th position in BWT is marked, Occ(1, 1, i) is index for genomic position in GI (e.g., GI[Occ(1,1,3)]=GI[2]=1)
- If i-th position in BWT is not marked, do LF_mapping until encounter marked position j, BWT[j] = 1, marked
- Pos(sp) = Number_of_LF_mappings + GI[Occ(1,1,j)]

```
LF_mapping(sp)
c = get_BWT_char(sp)
sp = F[c] + Occ(c, 1, sp)
```

```
Example: 1. LF_mapping(4)
    c = get_BWT_char(4) = T
    sp = F[T] + Occ(T, 1, 4) = 6 + 2 = 8 (not marked)
2. LF_mapping(8)
    c = get_BWT_char(8) = A
    sp = F[A] + Occ(A, 1, 8) = 1 + 2 = 3 (marked)
```

Pos(sp) = 2 + 1 = 3 (total of 2 LF_mappings and GI[Occ(1,1,3)])

Why LF_mapping works?

- 1 **\$ATATATTAG** 0
- 2 **AG\$ATATATT** 0
- 3 ATATATTAG\$ 1
- 4 ATATTAG\$AT 0
- 5 ATTAG\$ATAT 0
- 6 G\$ATATAT 0
- 7 TACSATATAT 1
- 8 TATATTAG\$A 0
- 9 TATTAG\$ATA 1

10**TTAG\$ATATA** 0

23456789

ATATATTAG

Why do we identify correctly the genomic position for unmarked BWT[i]?

- Given row M[i] starting with prefix P, we find the closest marked preceding genomic position
- Since the rows of M are the circular shifts of T\$, the last character of i-th row, L[i], precedes the first character F[i]
- Let L[i] = c and ri be the rank of the row M[i] among all rows ending with c. Then F[j] = c is the corresponding character to L[i] in T, where M[j] is the ri-th row of M starting with c
- Define LF_mapping as
 LF[i] = F[L[i]] + ri